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Phil. Trans. R. Soc. Lond. B 2000 **355**, 1007-1011
doi: 10.1098/rstb.2000.0637

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The dynamics of the cellular immune response to HIV infection: implications for vaccination

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Recent advances in measuring T-cell responses to viruses have led to new insights into how these T cells respond. In the acute infection there are massive CD8⁺ T-cell responses to both Epstein–Barr virus (EBV) and to human immunodeficiency virus (HIV). Many of these T cells are effector cells and only a minority appear to be capable of maintaining immunological memory. In persistent virus infections, high levels of antigen-specific effector cells persist. If virus does not persist, the effectors fade in number but memory is maintained and is primed to react rapidly to a new challenge. A vaccine that stimulates only T-cell responses may protect when these memory cells respond rapidly enough to generate high numbers of effectors before the infecting virus becomes established.

Keywords: HIV; Epstein–Barr virus; CD8⁺ T cells; memory; vaccination

1. INTRODUCTION

The study of the cellular immune response to infectious agents has been transformed by the recent introduction of major histocompatibility complex (MHC) tetramers (Altman *et al.* 1996; McMichael & O'Callaghan 1998). These are complexes of four MHC class I molecules containing the same epitope peptide linked by a biotin molecule to streptavidin. Such reagents enable direct visualization of antigen-specific T cells for the first time. In parallel, methods for intracellular cytokine staining of antigen-specific T cells have also been introduced (Butz & Bevan 1998; Pitcher *et al.* 1999). Use of these two technical advances has altered our view of how T cells respond to an invading pathogen. The biggest surprise was the magnitude of the CD8⁺ T-cell response to virus infections, ten to 100 times larger than many expected (Altman *et al.* 1996; Butz & Bevan 1998; Murali-Krishna *et al.* 1998; Tan *et al.* 1999). Nearly all the activated T cells seen in an acute infection are virus specific, very few are activated as bystanders. These findings permit a revision of how T cells respond, of their dynamics and what they do.

2. ACUTE HUMAN IMMUNODEFICIENCY VIRUS AND SIMIAN IMMUNODEFICIENCY VIRUS INFECTION

In acute human immunodeficiency virus (HIV) and simian immunodeficiency virus (SIV) infection, the virus replicates in an uncontrolled manner, reaching a peak of over 10⁶ particles per millilitre of blood in about three weeks. The CD8⁺ T-cell response peaks a few days later and at this time there is a fall in the viraemia. It has long been argued that these antigen-specific T cells are responsible for the reduction in virus load (Koup *et al.* 1994). Strong support for this view comes from studies in

macaques where CD8⁺ T cells were removed by antibody treatment *in vivo*; when challenged with SIV the viraemia was not controlled until the CD8⁺ T cells recovered from their suppression (Schmitz *et al.* 1999). The peak of the CD8⁺ T-cell response reaches 1–10% of all CD8⁺ T cells in the blood, both for HIV (Wilson *et al.* 2000) and SIV (Kuroda *et al.* 1999). It is not possible to measure the number of HIV-specific T cells in uninfected and unexposed people because the assays are not sensitive and accurate enough, but it is safe to estimate that the number is less than one in a million. Therefore it is likely that the T cells responding at the peak of the infection have divided over 15 times (figure 1). Essential to the use of the tetramer assay is knowledge of the dominant epitope peptide and it does appear that for many viruses the acute phase immune response is focused on very few epitopes out of the many that are theoretically possible (Chen *et al.* 2000). It looks as if the immunodominance is decided early on, for reasons that are still not clear (Chen *et al.* 2000).

HIV and SIV infect dendritic cells and CD4⁺ helper T cells, both of which are critical cells of the immune system. Their infection could affect the CD8⁺ T-cell response. Although the primary CD8⁺ T-cell response to HIV looks large, it appears to be smaller than the acute CD8⁺ T-cell response to Epstein–Barr virus (EBV) (Callan *et al.* 1998). Here CD8⁺ T cells specific for EBV have been found at levels as high as 44% of CD8⁺ T cells in the blood of patients with infectious mononucleosis; the range is between 5% and 45%. In acute infection with lymphocytic choriomeningitis virus in mice, similar high numbers were observed (Murali-Krishna *et al.* 1998).

In acute infectious mononucleosis, the expanded CD8⁺ T-cell population is capable of function (Callan *et al.* 2000). They can kill antigen-expressing target cells at about the level expected. They release cytokines (γ -interferon

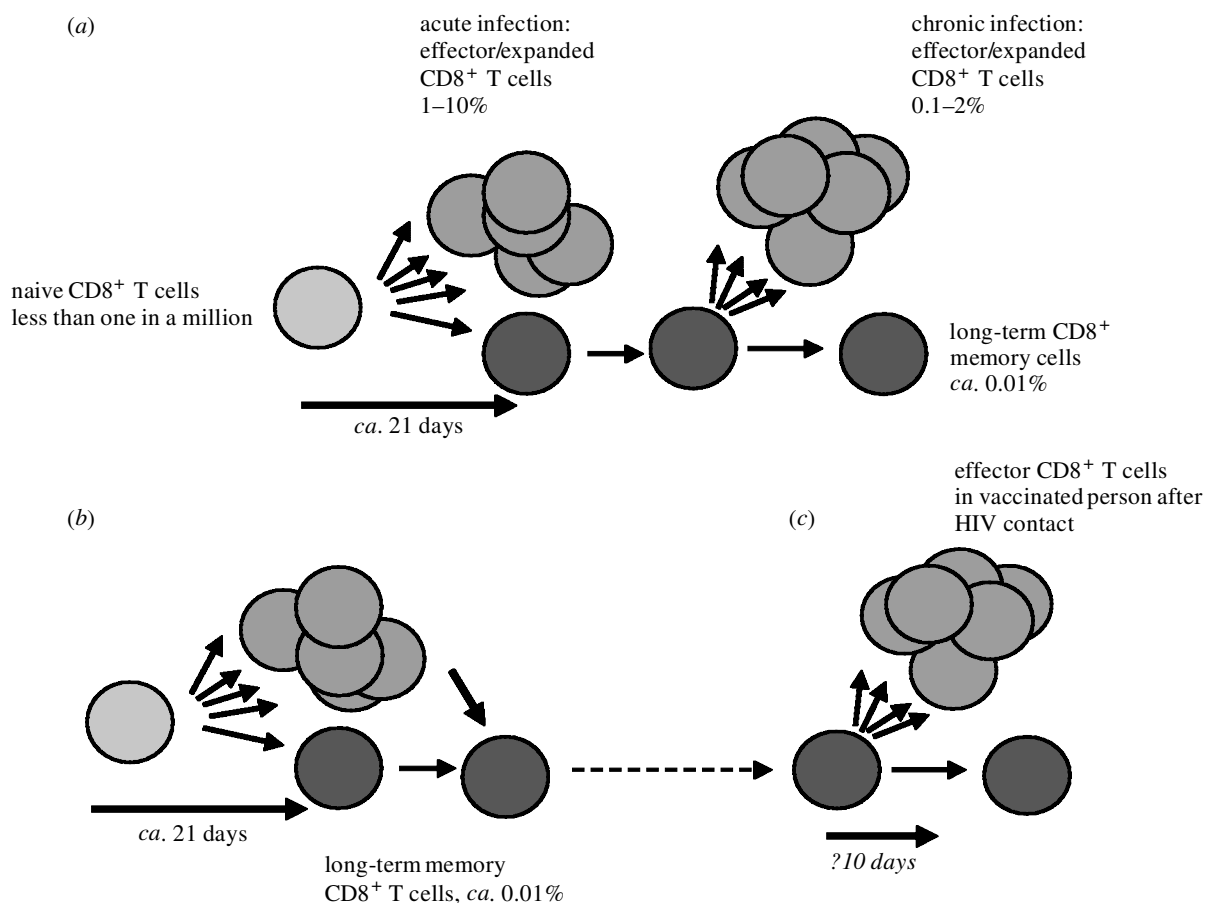


Figure 1. Possible dynamics of CD8⁺ T-cell response to (a) HIV, (b) a vaccine and (c) HIV after vaccination. In (a), the calculated numbers of naive HIV-specific T cells (estimate), expanded effectors (detected with tetramers) and long-term memory cells (detected by limiting dilution assay) are shown. In (b), the similar response to a non-persisting CD8⁺ T-cell stimulating vaccine is shown. (c) The response to HIV in a vaccinated person several months after vaccination. Figures are based on Hanke *et al.* (1999), Wilson *et al.* (2000) and Evans *et al.* (1999).

(IFN- γ), tumour necrosis factor- α (TNF- α) and macrophage inflammatory protein-1 β (MIP-1 β) rapidly on contact with antigen-presenting cells, although not all T cells can do this. However, on removal from the body, they die rapidly by apoptosis (Callan *et al.* 2000). Similarly, when stimulated by a non-persisting immunogen (DNA and recombinant modified vaccinia virus Ankara) the high CD8⁺ T-cell peak decays rapidly with a half-life in the blood of about seven days (Hanke *et al.* 1999). The expanded T cells appear therefore to be end-stage T cells, functioning but not capable of dividing further. Memory is maintained by a minor subset of T cells within the same clonal population that are able to divide further (Tan *et al.* 1999). The functional status of T cells in acute HIV infection is not yet known; preliminary experiments suggest some impairment of cytokine release and of perforin expression (V. Appay, unpublished data).

In acute HIV infection there is an early CD4⁺ T-cell response, but this has been hard to demonstrate, possibly because the acutely reacting T cells are infected and damaged by the HIV. However, Pitcher *et al.* (1999) have been able to demonstrate HIV-specific CD4⁺ T cells by using intracellular cytokine staining (after a few hours of exposure to antigen). The number found was substantial in patients with stable infection but less than the number

responding to cytomegalovirus, another persisting virus. Despite the clear presence of HIV-specific CD4⁺ T cells, it has been almost impossible to demonstrate them by the conventional antigen-stimulated proliferation assay; only a few long-term survivors show a response. However, Rosenberg *et al.* (1997) showed that it was possible to rescue this response by treating acutely infected persons very early with potent antiretroviral drugs. Thus there appears to be an early impairment of the CD4⁺ T-cell response that might affect the initiation of the CD8⁺ T-cell response. Nevertheless, the CD8⁺ T-cell response is substantial, mostly functional and important in reducing virus load. The suspicion remains, however, that it is not as effective as it might be and that this could result in higher virus loads at the set-point, once the acute phase stabilizes, than might otherwise be found.

3. CHRONIC HIV INFECTION

In chronic HIV infection, during the asymptomatic phase, there is a strong ongoing anti-HIV CD8⁺ T-cell response (Ogg *et al.* 1998). Often antigen-specific CD8⁺ T cells constitute over 1.0% of all CD8⁺ T cells when there is an identifiable, immunodominant T-cell response. Ogg *et al.* (1998) have shown that there is an

inverse correlation between the number of T cells specific for immunodominant gag and pol epitopes presented by HLA-A2 and the plasma virus RNA load. This is in a pseudo-equilibrium state and has been the subject of considerable discussion (see McMichael *et al.* 2000). In HTLV-1 infection, C. Bangham (personal communication) has shown that there is a positive correlation between virus load and CD8⁺ T-cell number, suggesting that the negative correlation reported by Ogg *et al.* (1998) might reflect suppression of the CD8⁺ T-cell response by the virus, presumably through the effects on HIV-specific CD4⁺ T cells. Mathematical modelling arguments support this model (D. Wodarz and M. Nowak, personal communication), as does the obvious finding that no virus means no detectable HIV-specific CD8⁺ T cells. We have suggested therefore that the relationship between virus load and cytotoxic T-lymphocyte (CTL) number is biphasic with a positive correlation at low virus load, but that above a certain virus level, HIV effectively suppresses the CTL response—most of the patients seen are in this second phase (D. Wodarz and M. Nowak, personal communication). This does not mean that the CTLs fail to suppress the virus—macaque experiments where the CD8⁺ T cells are removed *in vivo* in ongoing SIV infection, resulting in higher virus loads, support this view (Jin *et al.* 1999; Schmitz *et al.* 1999). However it does argue that the CD8⁺ T-cell response is not as effective as it might be in HIV infection.

This view is reinforced by the findings of Appay *et al.* (2000), who examined HIV-specific CD8⁺ T-cell function in samples from chronically infected persons. Whereas these T cells responded well to antigen stimulation to make IFN- γ , TNF- α and MIP-1 β , levels of perforin were very low. This was particularly striking when the HIV-specific T-cell responses were compared to cytomegalovirus-specific T cells in the same patients. The low perforin levels were found regardless of the virus load, even when undetectable as the result of aggressive antiretroviral drug therapy. Therefore it seems that perforin expression is impaired, perhaps a failure of CD8⁺ T-cell maturation due to poor CD4⁺ T-cell help (Zajac *et al.* 1998).

The high number of tetramer-staining cells in late HIV infection implies that they are surviving well or are actively replaced, from long-term memory T, after apoptosis. We favour the latter, though data to distinguish these possibilities are lacking. The number of CD8⁺ T cells specific for HIV remains high throughout the infection but the infection nevertheless progresses. Contributing factors to this are possible progressively greater impairment of CD8⁺ T-cell function and the selection of escape mutants. Appearance and selection of escape mutants has now been shown in several patients and also in macaques (Evans *et al.* 1999; Goulder *et al.* 1997; Phillips *et al.* 1991). The consequences are uncertain, but at the very least new CD8⁺ T-cell responses to alternative epitopes, i.e. new primary T-cell responses, must be required. Again, impaired CD4⁺ T-cell help must weaken these new responses and could contribute significantly to the gradual and harmful increase in virus load that occurs as the infection progresses.

4. IMPLICATIONS FOR VACCINES

Given the difficulty of current strategies to stimulate effective HIV-neutralizing antibodies in animals and humans (Connor *et al.* 1998), much attention has now been directed towards stimulating CD8⁺ T-cell responses. There are good data in animal models of other virus and parasite infections that CTL can protect against new infection (Brehm *et al.* 1997; Fu *et al.* 1999; Schneider *et al.* 1998). In HIV infection, extensive data from a small subset of African sex workers who are highly exposed to HIV but not infected, implies that their CTL response protects them from infection, as long as the CTL levels are maintained (Rowland-Jones *et al.* 1999). In macaque SIV infection models, data suggest that vaccine-induced CTL can protect or at least lower virus loads when infection occurs (Kent *et al.* 1998; Robinson *et al.* 1999).

How do CTLs protect against HIV infection? CTLs kill virus-infected cells and prevent them from producing new virus particles, but they cannot prevent infection, as they have no activity against naked virus. Furthermore, the kinetics of the antiviral CTL response shows that large numbers of activated effector T cells can be readily generated, but if antigen does not persist, they disappear rapidly (Hanke *et al.* 1999; figure 1b). Current vaccine constructs are designed not to persist, for safety reasons demanded by the regulatory authorities. Therefore it is likely that any protection will have to come from the long-term memory population.

Long-term memory T cells are probably best detected by limiting dilution assays, which demand multiple (about 12) divisions *in vitro* and maturation to effectors. The Elispot assay probably detects the same population in a situation where antigenic challenge occurred many months ago and antigen is no longer present, e.g. influenza infection, and detects very similar cell numbers to the limiting dilution assay (Lalvani *et al.* 1997). This indicates that long-term memory T cells can be activated to function (interferon secretion) within 6 h and before the cell divides (Lalvani *et al.* 1997). Thus in a person who was vaccinated several months ago by a CTL-inducing vaccine, there may not be instant killing of any HIV-infected cells that appear, but there is a response that differs substantially from that of the primary CTL response (figure 1c). The first advantage is in antigen-specific T-cell numbers: 10⁶ for an unprimed person and 10⁴ for a person with memory. This would give an advantage of seven divisions (2⁷), about seven days. Second, it has been shown that memory T cells respond much more rapidly than memory T cells and, as indicated above, can release cytokines almost immediately (Tanchot *et al.* 1998); this could give an advantage of a few more days. Thus, the vaccine-primed person should be able to respond much more rapidly to the new infection, although the magnitude of the response might be less if the virus load was well controlled. In addition, the ability of a vaccine to stimulate a good T-helper-cell response may give the primed person a further crucial advantage, ensuring that all the virus-specific T cells are functional and again speeding up the response. This has to be balanced for HIV infection with the possibility that these activated T cells will be good host cells for the virus to infect. Whether these advantages add up

enough to clear an incoming HIV infection remains to be seen, but there is a chance and at the least, the level of HIV at the set point should be lower, with a better prognosis.

5. CONCLUSIONS

The studies reviewed here give a kinetic view of HIV infection that is of crucial importance in understanding the natural response and how effective it is. This understanding is crucial for design of vaccines that can prime the T-cell response to be able to control infection and even clear it when the infecting challenge is low dose.

REFERENCES

- Altman, J. D., Moss, P., Goulder, P., Barouch, D. H., McHeyzer, W. M., Bell, J. I., McMichael, A. J. & Davis, M. M. 1996 Phenotypic analysis of antigen-specific T lymphocytes. *Science* **274**, 94–96. (Published erratum appears in *Science* 1998 **280**, 1821.)
- Appay, V. (and 15 others) 2000 HIV-specific CD8⁺ T cells produce antiviral cytokines but are impaired in cytolytic function. *J. Exp. Med.* (In the press.)
- Brehm, M. A., Bonneau, R. H., Knipe, D. M. & Tevethia, S. S. 1997 Immunization with a replication-deficient mutant of herpes simplex virus type 1 (HSV-1) induces a CD8⁺ cytotoxic T-lymphocyte response and confers a level of protection comparable to that of wild-type HSV-1. *J. Virol.* **71**, 3534–3544.
- Butz, E. A. & Bevan, M. J. 1998 Massive expansion of antigen-specific CD8⁺ T cells during an acute virus infection. *Immunity* **8**, 167–175.
- Callan, M. F., Tan, L., Annels, N., Ogg, G. S., Wilson, J. D., O'Callaghan, C. A., Steven, N., McMichael, A. J. & Rickinson, A. B. 1998 Direct visualization of antigen-specific CD8⁺ T cells during the primary immune response to Epstein-Barr virus *in vivo*. *J. Exp. Med.* **187**, 1395–1402.
- Callan, M. F., Fazou, C., Hatton, C. & McMichael, A. J. 2000 CD8⁺ T cell selection, function and death in the primary immune response *in vivo*. (Submitted.)
- Chen, W., Anton, L. C., Bennink, J. R. & Yewdell, J. W. 2000 Dissecting the multifactorial causes of immunodominance in class I-restricted T cell responses to viruses. *Immunity* **12**, 83–93.
- Connor, R. I. (and 10 others) 1998 Temporal analyses of virus replication, immune responses, and efficacy in rhesus macaques immunized with a live, attenuated simian immunodeficiency virus vaccine. *J. Virol.* **72**, 7501–7509.
- Evans, D. T. (and 16 others) 1999 Virus-specific cytotoxic T-lymphocyte responses select for amino-acid variation in simian immunodeficiency virus Env and Nef. *Nature Med.* **5**, 1270–1276.
- Fu, T. M., Guan, L., Friedman, A., Schofield, T. L., Ulmer, J. B., Liu, M. A. & Donnelly, J. J. 1999 Dose dependence of CTL precursor frequency induced by a DNA vaccine and correlation with protective immunity against influenza virus challenge. *J. Immunol.* **162**, 4163–4170.
- Goulder, P. J. (and 11 others) 1997 Late escape from an immunodominant cytotoxic T-lymphocyte response associated with progression to AIDS. *Nature Med.* **3**, 212–217.
- Hanke, T. (and 11 others) 1999 Effective induction of simian immunodeficiency virus-specific cytotoxic T lymphocytes in macaques by using a multiepitope gene and DNA prime-modified vaccinia virus Ankara boost vaccination regimen. *J. Virol.* **73**, 7524–7532.
- Jin, X. (and 13 others) 1999 Dramatic rise in plasma viremia after CD8⁺ T cell depletion in simian immunodeficiency virus-infected macaques. *J. Exp. Med.* **189**, 991–998.
- Kent, S. J., Zhao, A., Best, S. J., Chandler, J. D., Boyle, D. B. & Ramshaw, I. A. 1998 Enhanced T-cell immunogenicity and protective efficacy of a human immunodeficiency virus type 1 vaccine regimen consisting of consecutive priming with DNA and boosting with recombinant fowlpox virus. *J. Virol.* **72**, 10180–10188.
- Koup, R. A., Safrin, J. T., Cao, Y., Andrews, C. A., McLeod, G., Borkowsky, W., Farthing, C. & Ho, D. D. 1994 Temporal association of cellular immune responses with the initial control of viremia in primary human immunodeficiency virus type 1 syndrome. *J. Virol.* **68**, 4650–4655.
- Kuroda, M. J., Schmitz, J. E., Charini, W. A., Nickerson, C. E., Lifton, M. A., Lord, C. I., Forman, M. A. & Letvin, N. L. 1999 Emergence of CTL coincides with clearance of virus during primary simian immunodeficiency virus infection in rhesus monkeys. *J. Immunol.* **162**, 5127–5133.
- Lalvani, A., Brookes, R., Hambleton, S., Britton, W. J., Hill, A. V. & McMichael, A. J. 1997 Rapid effector function in CD8⁺ memory T cells. *J. Exp. Med.* **186**, 859–865.
- McMichael, A. J. & O'Callaghan, C. A. 1998 A new look at T cells. *J. Exp. Med.* **187**, 1367–1371.
- McMichael, A. J., Ogg, G., Wilson, J., Callan, M., Hambleton, S., Appay, V., Kelleher, A. & Rowland-Jones, S. 2000 Memory CD8⁺ T cells in HIV infection. *Phil. Trans. R. Soc. Lond. B* **355**, 363–367.
- Murali-Krishna, K., Altman, J. D., Suresh, M., Sourdive, D. J., Zajac, A. J., Miller, J. D., Slansky, J. & Ahmed, R. 1998 Counting antigen-specific CD8 T cells: a reevaluation of bystander activation during viral infection. *Immunity* **8**, 177–187.
- Ogg, G. S. (and 14 others) 1998 Quantitation of HIV-1-specific cytotoxic T lymphocytes and plasma load of viral RNA. *Science* **279**, 2103–2106.
- Phillips, R. E. (and 10 others) 1991 Human immunodeficiency virus genetic variation that can escape cytotoxic T cell recognition (see comments). *Nature* **354**, 453–459.
- Pitcher, C. J., Quittner, C., Peterson, D. M., Connors, M., Koup, R. A., Maino, V. C. & Picker, L. J. 1999 HIV-1-specific CD4⁺ T cells are detectable in most individuals with active HIV-1 infection, but decline with prolonged viral suppression (see comments). *Nature Med.* **5**, 518–525.
- Robinson, H. L. (and 16 others) 1999 Neutralizing antibody-independent containment of immunodeficiency virus challenges by DNA priming and recombinant pox virus booster immunizations (see comments). *Nature Med.* **5**, 526–534.
- Rosenberg, E. S., Billingsley, J. M., Caliendo, A. M., Boswell, S. L., Sax, P. E., Kalams, S. A. & Walker, B. D. 1997 Vigorous HIV-1-specific CD4⁺ T cell responses associated with control of viremia (see comments). *Science* **278**, 1447–1450.
- Rowland-Jones, S. L. (and 14 others) 1999 Broadly cross-reactive HIV-specific cytotoxic T-lymphocytes in highly-exposed persistently seronegative donors. *Immunol. Lett.* **66**, 9–14.
- Schmitz, J. E. (and 15 others) 1999 Control of viremia in simian immunodeficiency virus infection by CD8⁺ lymphocytes. *Science* **283**, 857–860.
- Schneider, J., Gilbert, S. C., Blanchard, T. J., Hanke, T., Robson, K. J., Hannan, C. M., Becker, M., Sinden, R., Smith, G. L. & Hill, A. V. 1998 Enhanced immunogenicity for CD8⁺ T cell induction and complete protective efficacy of malaria DNA vaccination by boosting with modified vaccinia virus Ankara. *Nature Med.* **4**, 397–402.
- Tan, L. C., Gudgeon, N., Annels, N. E., Hansasuta, P., O'Callaghan, C. A., Rowland-Jones, S., McMichael, A. J.,

Rickinson, A. B. & Callan, M. F. 1999 A re-evaluation of the frequency of CD8⁺ T cells specific for EBV in healthy virus carriers. *J. Immunol.* **162**, 1827–1835.

Tanchot, C., Guillaume, S., Delon, J., Bourgeois, C., Franzke, A., Sarukhan, A., Trautmann, A. & Rocha, B. 1998 Modifications of CD8⁺ T cell function during *in vivo* memory or tolerance induction. *Immunity* **8**, 581–590.

Wilson, J. D. (and 10 others) 2000 Direct visualization of HIV-1 specific cytotoxic T lymphocytes during primary infection. *AIDS* **14**, 225–233.

Zajac, A. J., Blattman, J. N., Murali-Krishna, K., Sourdive, D. J., Suresh, M., Altman, J. D. & Ahmed, R. 1998 Viral immune evasion due to persistence of activated T cells without effector function (see comments). *J. Exp. Med.* **188**, 2205–2213.